Accepted: 16.04.2016

## **Original Article**

## *Helianthus annuus* Leaf Ameliorates Postprandial Hyperglycaemia by Inhibiting Carbohydrate Hydrolyzing Enzymes Associated with Type-2 Diabetes

Oluwafemi Adeleke Ojo<sup>\*1</sup>, Adebola Busola Ojo<sup>2</sup>, Basiru Ajiboye<sup>1</sup>, Israel Olayide<sup>1</sup>, Adewale Fadaka<sup>1</sup>

Received: 03. 03 2016

## ABSTRACT

**Background:** Diabetes mellitus is a chronic disease. Decreasing postprandial hyperglycemia by retarding glucose absorption through inhibiting carbohydrates digesting enzymes ( $\alpha$ -amylase and  $\alpha$ -glucosidase) is one of many approaches used for the management of this disease. This study was aimed at evaluating the normoglycaemic potential of *Helianthus annuus* leaf.

**Methods:** The effect of the *in vitro* inhibitory of different extracts (acetone, ethyl acetate and hexane) of the plant was assessed on the activities of diabetes-related enzymes ( $\alpha$ -amylase and  $\alpha$ -glucosidase).

**Results:** The hexane extract of *H. annuus* leaf displayed the best inhibitory activity against  $\alpha$ -amylase and  $\alpha$ -glucosidase as indicated by the IC<sub>50</sub> values (3.92 ± 0.02 mg mL<sup>-1</sup>) and (3.29 ± 0.12 mg mL<sup>-1</sup>), respectively. Lineweaver-Burk plot of inhibition of  $\alpha$ -amylase and  $\alpha$ -glucosidase by this extract showed that it was competitive and non-competitive mode, respectively.

**Conclusion:** *H. annuus* leaf possesses hypoglycaemic potential which may be due to the inhibition of pancreatic  $\alpha$ -amylase and intestinal  $\alpha$ -glucosidase.

**Keywords:** α-Amylase, α-Glucosidase, *Helianthus Annuus*, Postprandial Hyperglycaemia.

#### **INTRODUCTION**

The prevalence of type-2 diabetes mellitus is increasing at an alarming rate such that is currently being estimated to be responsible for 90-95% of all diabetes cases worldwide [1]. This occurs as a result of lifestyle and socio-economic changes mainly characterized by lower physical activity and higher intake of fat and saturated carbohydrate containing diets among many other factors [2]. Hyperglycemia is a hallmark of type-2 diabetes mellitus and plays a vital role in most of the pathogenic features of the disease. This condition prevails when there is decreased insulin sensitivity or decreased insulin secretion from pancreatic  $\beta$ -cells, which can inhibit insulin secretion from the pancreas and reduce insulin mediated glucose uptake in peripheral tissues [3, 4]. All diabetic complications (nephropathy, neuropathy, microangiopathy, macroangiopathy, retinopathy, and cataract) are strongly linked to hyperglycemia. Therefore, the need for the improved treatment of hyperglycemia, type 2 diabetes mellitus related risk factors and the long term degenerative disorders so as to drastically lower the risk of both micro and macro vascular

### IJT 2016 (5): 17-22

complications. An important strategy to control hyperglycemia is through the inhibition of key carbohydrates digesting enzymes such as  $\alpha$ amylase and  $\alpha$ -glucosidase, which also play a vital role in preventing diabetic complications. The inhibitors of these enzymes delay the digestion of carbohydrates reduce the rate of glucose absorption from the small intestinal tract, therefore, reducing postprandial blood glucose level. Thus, inhibition of  $\alpha$ -amylase and  $\alpha$ glucosidase is a key in the management and treatment of hyperglycemia and type 2 diabetes mellitus [3].

Helianthus annuus L. is a folk remedy for bronchiectasis, bronchitis, carbuncles, catarrh, cold, colic, cough, diarrhea, dysentery, dysuria, epistaxis, eyes, fever, flu, fractures, inflammations. laryngitis, malaria. lungs, menorrhagia, pleuritis, rheumatism, scorpion stings, snakebite, splenitis, urogenital ailments, whitlow, and wounds [5]. Anti-hyperglycaemic effects of these plants are due to their capability to improve the function of pancreatic tissues by causing an increase in insulin output or inhibit the intestinal absorption of glucose or the facilitation

<sup>1.</sup> Department of Biochemistry, Afe Babalola University, Ado-Ekiti, Nigeria.

<sup>2.</sup> Department of Biochemistry, Ekiti State University, Ado-Ekiti, Nigeria.

<sup>\*</sup> Corresponding Author: E-mail: oluwafemiadeleke08@gmail.com

of metabolites in insulin dependent processes [6]. *H. annuus* leaves are extensively used for whooping cough, asthma, anti- malaria, insect bites, snake bites, fevers and lung problems. Elsewhere, the roots are used for treatment of diabetics [7]. Despite the usage of *H. annuus* leaf in the management of diabetes mellitus, there is no scientific report on the antidiabetic potential of this plant.

Therefore, the present study aimed to assess the hypoglycaemic potential of *H. annuus* leaf, through the evaluation of inhibitory potential of its extracts on key enzymes linked to diabetes mellitus ( $\alpha$ -amylase and  $\alpha$ -glucosidase).

## MATERIALS AND METHODS

#### Sample Collection

Fresh Leaves of *H. annuus* were collected at a Ijadu farm in the suburbs of Ado Ekiti, Nigeria. The plant was identified and authenticated by a plant scientist in the Department of Plant Science, Ekiti State University, Ado-Ekiti, Nigeria and a voucher specimen number was deposited accordingly at the herbarium of the Department of Plant Science, Ekiti State University, Ado-Ekiti, Nigeria.

#### Chemicals and Reagents

 $\alpha$ -glucosidase **Saccharomyces** from porcine cerevisiae, pancreatic amylase, pnitrophenyl-a-D-glucopyranoside (pNPG), *p*nitrophenol, gallic acid. potassium and ferricyanide were obtained from Sigma-Aldrich, Germany. Starch, dinitrosalicylic acid (DNS), maltose, ethanol, ethyl acetate, trichloroacetic acid reagents were obtained from Merck Chemical Company, Germany.

#### **Preparation of Plant Extracts**

The powdered leaves were divided into three portions of 50 g each and these were extracted with acetone, ethyl acetate and hexane. They were all left to steep in covered conical flasks for 24 h, the flasks were shaken and kept still to allow the plant material settle at the bottom of the flask. The resulting infusions were decanted, filtered and evaporated in a rotary evaporator. Dried extracts were weighed and dissolved in dimethylsulphoxide (DMSO) to yield a stock solution from which lower concentrations were prepared. % yield=  $\frac{Weight of the dry extract}{Weight of powdered leaves} \times 100\%$ 

## Preliminary Phytochemical Screening

*Helianthus annuus* L., was subjected to qualitative chemical screening to identify the various major classes of active chemical constituents, namely tannins, steroid, terpenoids, saponins, cardiac glycosides, flavonoids, and alkaloid [8, 9].

## Determination of *a*-Amylase Inhibitory Activity

The  $\alpha$ -amylase inhibitory activity was determined [10], with slight modifications. A volume of 250 µL of ethyl acetate fraction at different concentrations (1-5 mg mL<sup>-1</sup>) was incubated with 500 µL of porcine pancreatic amylase (2 U mL<sup>-1</sup>) in 100 mmol L-1 phosphate buffer (pH 6.8) at 37 °C for 20 min. Two hundred and fifty µL of 1 % starch dissolved in 100 mmol  $L^{-1}$  phosphate buffer (pH 6.8) was then added to the reaction mixture and incubated at 37 °C for 1 h. One mL of DNS color reagent was then added and boiled for 10 min. The absorbance of the resulting mixture was measured at 540 nm and the inhibitory activity was expressed as percentage of a control sample without inhibitors. All assays were carried out in triplicate.

 $\propto$  – amylase inhibition (%) =

- × 100

A540control – A540sample

A540control

## Mode of a-Amylase Inhibition

The mode of inhibition of  $\alpha$ -amylase by the leaf extract was conducted using the most potent extract according to a modified method [11]. Briefly, 250  $\mu$ L of the (2.5 mg mL<sup>-1</sup>) extract was pre-incubated with 250  $\mu$ L of  $\alpha$ -amylase solution for 10 min at 25 °C in one set of tubes. In another set of tubes,  $\alpha$ -amylase was pre-incubated with 250 µL of phosphate buffer (pH 6.9). Two hundred and fifty microliter of starch solution at increasing concentrations  $(0.3-5.0 \text{ mg mL}^{-1})$  was added to both sets of reaction mixtures to start the reaction. The mixture was then incubated for 10 min at 25 °C and then boiled for 5 min after addition of 500 µL of DNS to stop the reaction. The amount of reducing sugars released was spectrophotometrically determined using а maltose standard curve and converted to reaction velocities. A double reciprocal (Lineweaver-Burk)

plot (1/v versus 1/[S]) where, v is reaction velocity and [S] is substrate concentration was plotted to determine the mode of inhibition.

#### Determination of a-Glucosidase Inhibitory Activity

The  $\alpha$ -glucosidase inhibitory activity was determined [12], with slight modifications. Briefly, 250 µL of ethyl acetate fraction, at different concentrations (1–5 mg mL<sup>-1</sup>), was incubated with 500 µL of 1.0 U mL<sup>-1</sup>  $\alpha$ -glucosidase solution in 100 mmol L<sup>-1</sup> phosphate buffer (pH 6.8) at 37 °C for 15 min. Thereafter, 250 µL of pNPG solution (5 mmol L<sup>-1</sup>) in 100 mmol L<sup>-1</sup> phosphate buffer (pH 6.8) was added and the mixture was further incubated at 37 °C for 20 min. The absorbance of the released *p*-nitro phenol was measured at 405 nm and the inhibitory activity was expressed as percentage of a control sample without inhibitors.

# $\frac{\alpha - \text{glucosidase inhibition (\%)} = \frac{\text{A405control} - \text{A405sample}}{\text{A405control}} \times 100$

#### Mode of a-Glucosidase Inhibition

The mode of inhibition of  $\alpha$ -glucosidase by the extracts was determined using the extract with the lowest IC<sub>50</sub> according to a modified method [11]. Briefly, 50  $\mu$ L of the (2.5 mg mL<sup>-1</sup>) extract was pre-incubated with 100  $\mu$ L of  $\alpha$ -glucosidase solution for 10 min at 25 °C in one set of tubes. In another set of tubes, a-glucosidase was preincubated with 50 µL of phosphate buffer (pH 6.9). 50 µL of pNPG at increasing concentrations  $(0.63-2.0 \text{ mg mL}^{-1})$  was added to both sets of reaction mixtures to start the reaction. The mixture was then incubated for 10 min at 25 °C and 500 µL of Na<sub>2</sub>CO<sub>3</sub> was added to stop the reaction. The amount of reducing sugars released was determined spectrophotometrically using a para-nitrophenol standard curve and converted to reaction velocities. double А reciprocal (Lineweaver-Burk) plot (1/v versus 1/[S]) where v is reaction velocity and [S] is substrate concentration was plotted to determine the mode of inhibition.

#### Statistical Analysis

Statistical analysis was performed using GraphPad Prism 5 statistical package (GraphPad Software, USA). All the results were expressed as Mean  $\pm$  SEM for triplicate determinations.

#### RESULTS

Phytochemical screening of the extracts of *H. annuus* L. showed the presence of various chemical constituents, mainly alkaloids, saponins, polysaccharides, flavonoids, polyphenols. The results obtained were comparable and satisfied the standard literature.

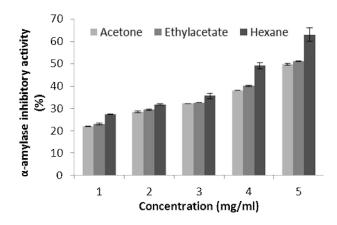
Figure 1 shows the result of the  $\alpha$ -amylase inhibitory potential of different extracts of H. annuus leaf. At the lowest concentration tested (1 mg mL<sup>-1</sup>), there were no significant differences among all the extracts. At higher concentrations, there were no significant differences between the acetone and ethyl acetate extracts. However, the percentage inhibition of the enzyme by hexane significantly higher (P < 0.05)extract was compared to other extracts. This is corroborated by the lowest IC<sub>50</sub> value  $(3.92 \pm 0.02 \text{ mg mL}^{-1})$ exhibited by the hexane extract compared to other extracts (Table 1). The Lineweaver-Burk plot showed that hexane extract of H. annuus leaf displayed competitive inhibition of the enzyme,  $\alpha$ amylase (Figure. 2).

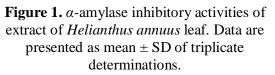
Figure 3 shows the inhibitory effect of different extracts of *H. annuus* leaf on  $\alpha$ -glucosidase. Across all the concentrations, hexane extract displayed significantly higher (*P*<0.05) percentage inhibition of the enzyme than acetone and ethyl acetate extracts while there was no significant difference between the inhibitions by acetone and ethyl acetate. The higher percentage inhibition exhibited by the hexane extract resulted in its lowest IC<sub>50</sub> (3.29 ± 0.12 mg mL<sup>-1</sup>). The hexane extract of *Helianthus annuus* leaf also inhibited  $\alpha$ -glucosidase in a mixed non-competitive manner as shown in (Figure. 4).

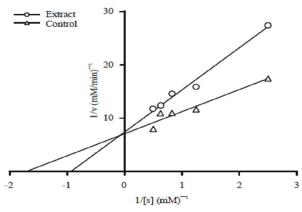
**Table 1.** IC<sub>50</sub> values for the inhibition of  $\alpha$ amylase and  $\alpha$ -glucosidase by *Helianthus annuus* leaf

Extracts	IC <sub>50</sub> (mg/ml)	
	$\alpha$ -amylase	$\alpha$ -glucosidase
Acetone	$5.42\pm0.22^{\rm a}$	$4.43\pm0.52^{\rm a}$
Ethyl acetate	$5.20\pm0.18^{\rm a}$	$4.05\pm0.68^{\rm a}$
Hexane	$3.92\pm0.02^{\rm b}$	$3.29\pm0.12^{b}$

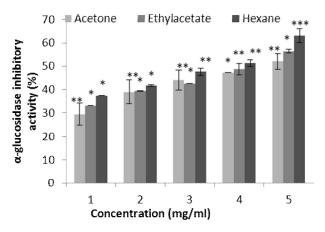
Data are presented as mean  $\pm$  SD of duplicate determinations. Different letters (a,b) over the bars for a given concentration for each extract indicate a significant difference from each other (Duncan multiple range post hoc test (*P*<0.05).



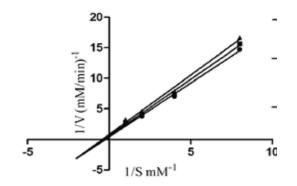




**Figure 2.** Mode of inhibition of  $\alpha$ -amylase by hexane extract of leaf of *Helianthus annuus*.



**Figure 3.**  $\alpha$ -glucosidase inhibitory activities of ethanol extract of *Helianthus annuus* leaf. Data are presented as mean  $\pm$  SD of triplicate determinations. (\*-\*\*\*) Different symbol over the bars for a given concentration for each extract indicate a significant difference from each other (Duncan multiple range *post hoc* test, *P* < 0.05).



**Figure 4.** Mode of inhibition of  $\alpha$ -glucosidase by hexane extract of leaf of *Helianthus annuus*.

#### DISCUSSION

The anti-diabetic effect of *H. annuus* leaf of different extracts (acetone, ethyl acetate and hexane) was investigated *in vitro*.

Although, pharmacological activities of medicinal plants are as a result of their chemical components, this is what informed the phytochemical screening of the three extracts of H. annuus leaf used in this study. The presence of only alkaloids, cardiac glycosides, tannins and saponins was detected in the hexane extract. Alkaloids have been shown to display wide spectrum antidiabetic potentials by inhibiting diabetes-related enzymes like a-glucosidase and preventing protein glycation [13, 14]. Tannins on the other hand, exhibits insulin-mimetic effect by improving the sensitivity of insulin receptors [15, 16] while cardiac glycosides prevent diabetesrelated complications by inhibiting  $Na^+/K^+$  pump, thereby improving cardiac output and preventing cardiovascular diseases [17]. Saponins on the other hand have been reported to having insulin sensitization and anti-hyperlipidemic effects in diabetic state [18, 19].

 $\alpha$ -Glucosidase and  $\alpha$ -amylase inhibitors have been useful as oral hypoglycemic drugs for the control of hyperglycemia in patients with type 2 diabetes. Inhibitions of these enzymes delay carbohydrate digestion and overall prolong the digestion time causing a reduction in the rate of glucose absorption and consequently reducing postprandial plasma glucose [20]. However, all the extracts of *H. annuus* leaf inhibited both  $\alpha$ amylase and  $\alpha$ -glucosidase. The highest percentage inhibition of the  $\alpha$ -amylase by the hexane extract resulted in its low IC<sub>50</sub> value (3.92  $\pm 0.02 \text{ mg mL}^{-1}$ ) which implies that it is the most potent inhibitor of the enzyme out of the three extracts. However, it is undesirable of an

DOI: 10.29252/arakmu.10.5.17

antidiabetic agent to be a strong inhibitor of  $\alpha$ amylase so as to prevent some of the drawbacks of synthetic drugs which might be due to excessive inhibition of the enzyme [21]. The competitive inhibition of  $\alpha$ -amylase by the hexane extract of *H. annuus* leaf suggests that the active inhibitory component(s) of the extract are structurally similar to the normal substrate of the enzyme. However, it binds reversibly to the active site of the enzyme and occupies it in a mutually exclusive manner with the substrate [10].

However, for  $\alpha$ -glucosidase inhibition, the strong inhibition of the enzyme displayed by the extracts as shown by the  $IC_{50}$  (Table 1) values  $(3.29 \pm 0.12 \text{ mg mL}^{-1})$  suggests that hexane extract is a potent  $\alpha$ -glucosidase inhibitor. This is in conformity with previous reports that antidiabetic agents from plants are strong inhibitors of  $\alpha$ -glucosidase [21, 22]. This implies that hexane extract of H. annuus leaf offer better pharmacological effect than the common synthetic drugs. Further study to ascertain the mode of inhibition  $\alpha$ -glucosidase (Figure 4) by the hexane extract of H. annuus leaf showed the extracts inhibited the enzyme in a mixed non-competitive inhibition. The mixed non-competitive mode of inhibition obtained from the Lineweaver-Burk plot shows that the active components in the hexane extract do not compete with the substrate for binding to the active site rather the inhibitors bind to a separate site on the enzyme to retard the conversion of disaccharides to monosaccharides [23].

#### CONCLUSION

*H. annuus* leaf possesses anti-diabetic potential in Wistar rats and the possible mechanism of action in the inhibition of pancreatic  $\alpha$ -amylase and  $\alpha$ -glucosidase, thereby slowing down the absorption of carbohydrates and preventing hyperglycaemia. The hypoglycaemic activity of this plant may also be due to the presence of alkaloids, tannins and saponins.

#### ACKNOWLEDGMENT

The authors wish to acknowledge the assistance given by Department of Biochemistry, Afe Babalola, Ado-Ekiti. The authors declare no conflict of interest.

#### REFERENCES

1. Wild S, Roglic G, Green A, Sicree R, King H. Global prevalence of diabetes estimates for the

year 2000 and projections for 2030. Diabetes care 2004;27(5):1047-53.

- 2. International Diabetes Federation (IDF). Diabetes Atlas. 5 <sup>th</sup> ed. Brussels: Belgium, 2011. P.14-5.
- 3. Ali MS, Jahangir M, ul Hussan SS, Choudhary MI. Inhibition of  $\alpha$ -glucosidase by oleanolic acid and its synthetic derivatives. Phytochemistry 2002;60(3):295-9.
- Cai H, Mcneilly AS, Luttrell LM, Martin B. Endocrine function in aging. Int J Endocrinol 2012; 2012:872478-81.
- Duke J, Wain K. Medicinal plants of the world. Computer index with more than 85000 entries. Encyclopaedia Americana, Grolier 1981;3:25-6.
- Ngondi JL, Fossouo Z, Djiotsa EJ, Oben J. Glycaemic variations after administration of Irvingia gabonensis seeds fractions in normoglycemic rats. Afr J Tradit Complement Altern Med 2006;3(4):94-101.
- Saini S, Sharma S. Antidiabetic effect of Helianthus annuus L, seeds ethanolic extract in streptozotocinnicotinamide induced type 2 diabetes mellitus. Int J Pharm Pharm Sci 2013;5(2):382-7.
- Sofowora A. Screening plants for bioactive agents. In: Medicinal plants and traditional medicinal in Africa. Nigeria: Ibadan;1993.
- 9. Evans W. Trease and Evans pharmacognosy. WB Saunders and Company, London. 2002.
- Shai LJ, Masoko P, Mokgotho MP, Magano SR, Mogale A, Boaduo N, et al. Yeast alpha glucosidase inhibitory and antioxidant activities of six medicinal plants collected in Phalaborwa, South Africa. Afr J Bot 2010;76(3):465-70.
- Ali H, Houghton P, Soumyanath A. α-Amylase inhibitory activity of some Malaysian plants used to treat diabetes; with particular reference to Phyllanthus amarus. J Ethnopharmacol 2006;107(3):449-55.
- 12. Ademiluyi AO, Oboh G. Soybean phenolic-rich extracts inhibit key-enzymes linked to type 2 diabetes ( $\alpha$ -amylase and  $\alpha$ -glucosidase) and hypertension (angiotensin I converting enzyme) in vitro. Exp Toxicol Pathol 2013;65(3):305-9.
- 13. Choudhary MI, Adhikari A, Rasheed S, Marasini BP, Hussain N, Kaleem WA. Cyclopeptide alkaloids of Ziziphus oxyphylla Edgw as novel inhibitors of  $\alpha$ -glucosidase enzyme and protein glycation. Phytochem Lett 2011;4(4):404-6.
- Ojo OA, Oloyede OI, Olarewaju OI, Ojo AB, Ajiboye BO, Onikanni SA. Toxicity studies of the crude aqueous leaves extracts of *Ocimum* gratissimum in albino rats. IOSR J Environ Sci Toxicol Food Technol 2013; 6(4) :34-9.
- 15. Huang TH, Peng G, Kota BP, Li GQ, Yamahara J, Roufogalis BD, et al. Anti-diabetic action of

Punica granatum flower extract: activation of PPAR- $\gamma$  and identification of an active component. Toxicol Appl Pharmacol 2005;207(2):160-9.

- 16. Ajiboye BO, Ibukun EO, Edobor G, Ojo OA, Onikanni SA. Qualitative and Quantitative analysis of phytochemicals in *Senecio biafrae* leaf. Int J Inv Pharm Sci 2013; 1(5): 428-32.
- 17. Krishnaiah D, Devi T, Bono A, Sarbatly R. Studies on phytochemical constituents of six Malaysian medicinal plants. JMPR 2009;3(2):067-72.
- Lee K-T, Jung TW, Lee H-J, Kim S-G, Shin Y-S, Whang W-K. The antidiabetic effect of ginsenoside Rb2 via activation of AMPK. Arch Pharm Res 2011;34(7):1201-8.
- 19. Eu CHA, Lim WYA, Ton SH, bin Abdul Kadir K. Glycyrrhizic acid improved lipoprotein lipase expression, insulin sensitivity, serum lipid and lipid deposition in high-fat diet-induced obese rats. Lipids Health Dis 2010;9(1):1-9.

- Kimura Y, Araki Y, Takenaka A, Igarashi K. Protective effects of dietary nasunin on paraquatinduced oxidative stress in rats. Biosci Biotechnol Biochem 1999;63(5):799-804.
- 21. Kwon YI, Apostolidis E, Shetty K. Inhibitory potential of wine and tea against  $\alpha$ -Amylase and  $\alpha$ -Glucosidase for management of hyperglycemia linked to type 2 diabetes. J Food Biochem 2008;32(1):15-31.
- 22. Kazeem MI, Ogungbe SM, Saibu GM, Aboyade OM. In vitro study on the hypoglycemic potential of Nicotiana tabacum leaf extracts. Bangladesh J Pharmacol 2014;9(2):140-5.
- Mogale M, Lebelo SL, Thovhogi N, De Freitas A, Shai L. α-Amylase and α-glucosidase inhibitory effects of Sclerocarya birrea [(A. Rich.) Hochst.] subspecies caffra (Sond) Kokwaro (Anacardiaceae) stem-bark extracts. AJB 2011;10(66):15033-9.